

Chap 5, 19

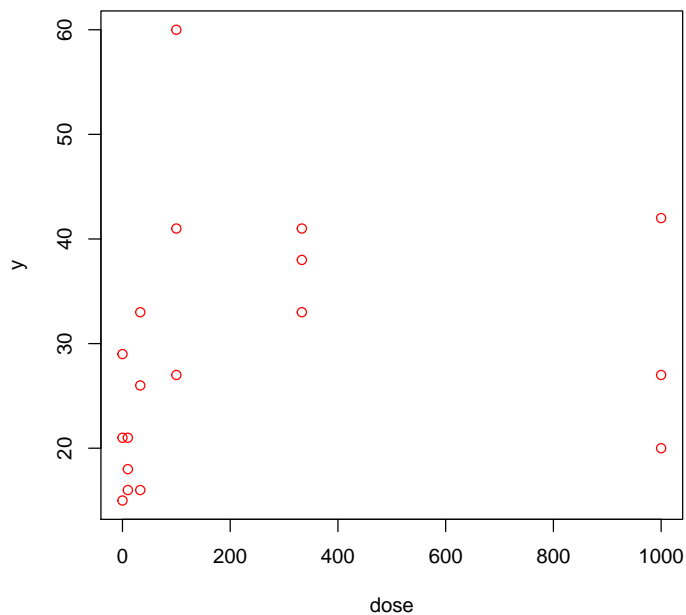
(a) It's easier to see the distribution of the points when the x-axis of these points are evenly distributed

(b) The p-values are 0.1,  $<1e-5$ ,  $<1e-5$  for the base, log10, and factor models, so the first is not significant, but significant for the other two.

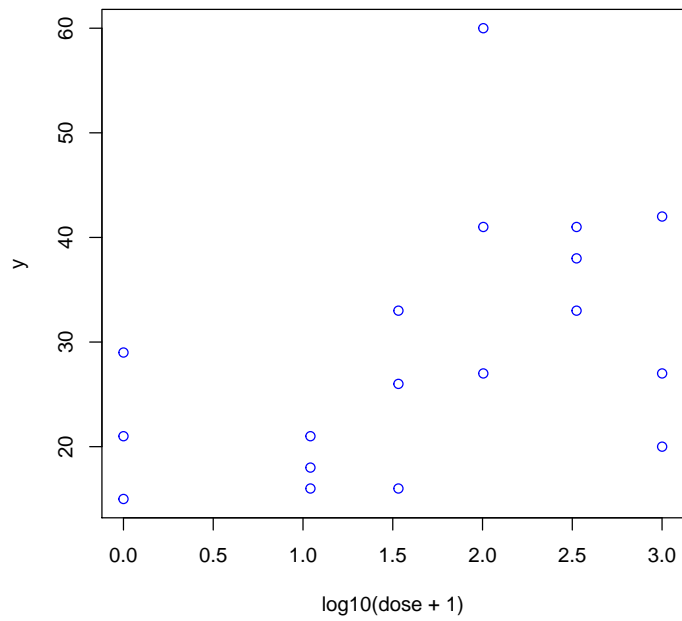
(c) Compare Dev/DF with 2 (like the critical value 1.96), the "factor" model (Dev/DF = 2.8) is less lack of fit. The "dose" model (Dev/DF = 4.7) is most lack of fit.

Model	$-2 \log \Lambda$	DF	p-value	Residual Dev	Residual DF	Dev/DF
dose	2.55	1	0.11	75.8	16	4.7
log10(dose+1)	18.73	1	1.5e-5	59.6	16	3.7
factor(dose)	44.86	5	1.5e-8	33.4	12	2.8

```
library(aod)
data("salmonella")
dose = salmonella$dose
y = salmonella$y
plot(dose, y, col="red")
```



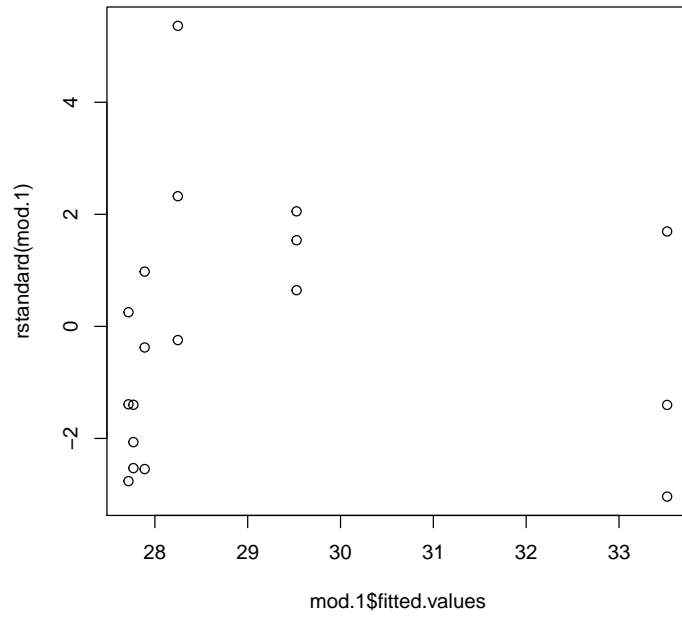
```
plot(log10(dose+1), y, col="blue")
```



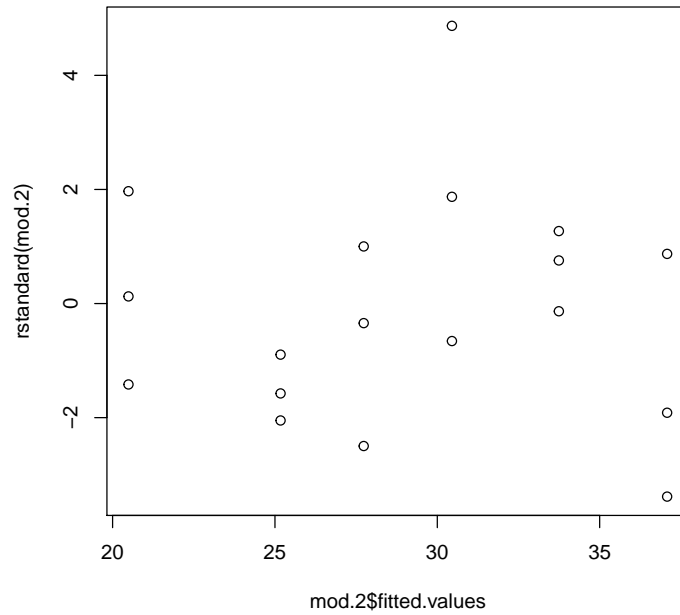
```
mod.1 <- glm( y ~ dose, family = poisson())  
mod.2 <- glm( y ~ log10(dose+1), family = poisson())  
mod.3 <- glm( y ~ as.factor(dose), family = poisson())  
A3 = anova(mod.3, test = "Chisq")
```

(d)

```
plot(mod.1$fitted.values, rstandard(mod.1))
```

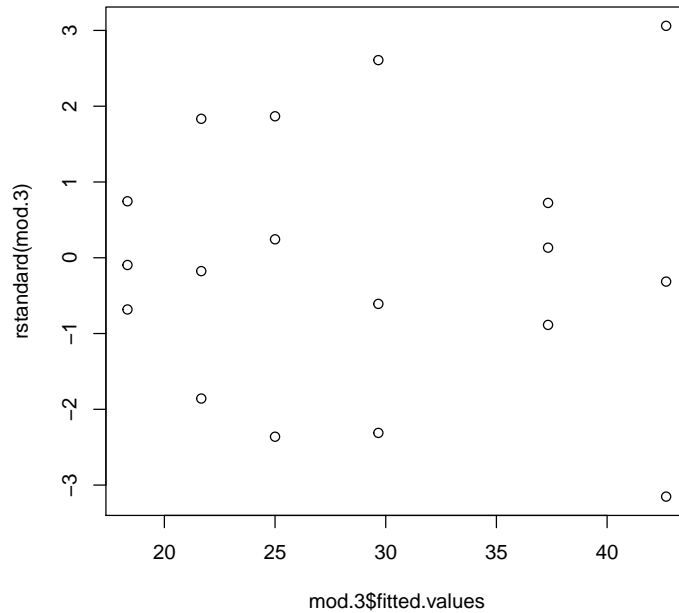


```
plot(mod.2$fitted.values, rstandard(mod.2))
```



(e) The mean residual is now 0 everywhere, and so the extreme residuals are not so large. There are still two that lie beyond  $\pm 3$  and three more beyond  $\pm 2$ , which is too many for a sample of size 18. There is some kind of overdispersion relative to a Poisson model.

```
plot(mod.3$fitted.values, rstandard(mod.3))
```



Chap 5, 35

(a) Negative-binomial has smaller deviance residual, so it is better than Quasipoisson.

```

library(MASS)
library(car)
mod.nb <- glm.nb(y ~ as.factor(dose))
mod.qp <- glm(y ~ as.factor(dose), family = quasipoisson(link = "log"))
print(as.matrix(anova(mod.qp, mod.nb)),quote = "FALSE")
  Resid. Df Resid. Dev Df Deviance
1      12  33.49596 NA      NA
2      12  17.27234  0 16.22362

res.sq <- residuals ( object = mod.3 , type = "response")^2
set1 <- data.frame ( res.sq , mu.hat = mod.3$fitted.values )
fit.lin <- lm( formula = res.sq ~ mu.hat , data = set1 )
fit.quad <- lm( formula = res.sq ~ mu.hat + I(mu.hat ^2), data = set1 )
plot ( x = set1$mu.hat , y = set1$res.sq , xlab = "Predicted count" , ylab = "Squared R
curve ( expr = predict ( object = fit.lin , newdata =
      data.frame (mu.hat = x ) , type = "response" ) , col = "blue" , add
      = TRUE , lty = "solid" )
curve (expr = predict ( object = fit.quad , newdata =
      data.frame ( mu.hat = x ) , type = "response" ) , col = "red" , add =
      TRUE , lty = "dashed" )

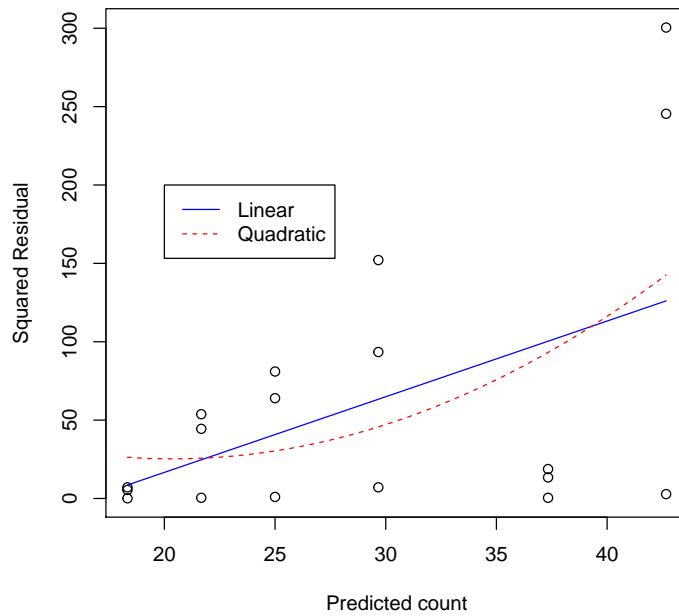
```

```

legend ( x = 20 , y = 200 , legend = c ("Linear" , "Quadratic") , col
       = c ("blue", "red") , lty = c ("solid" , "dashed") , bty = "n ")
print(as.matrix(anova(mod.qp, test="Chisq")), quote = "FALSE")

```

	Df	Deviance	Resid. Df	Resid. Dev	Pr(>Chi)
NULL	NA	NA	17	78.35758	NA
as.factor(dose)	5	44.86162	12	33.49596	0.006431595

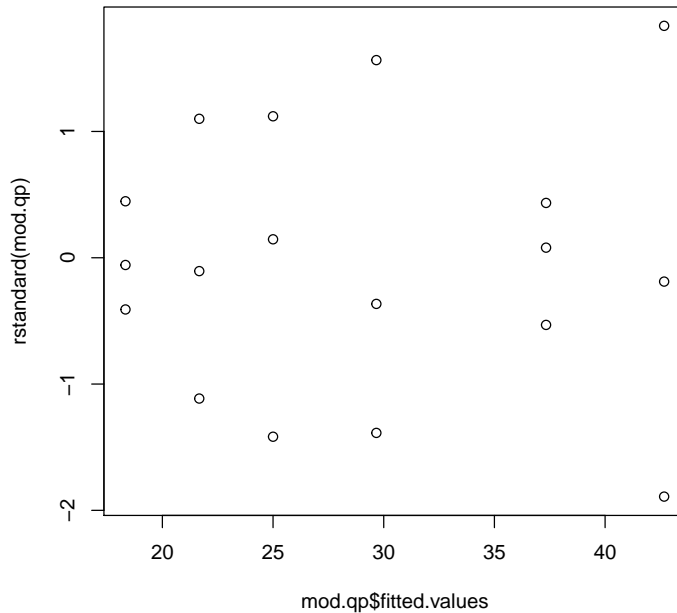


- (b) The p-value is 0.006432, which indicates the dose has an significant effect.  
(c)

```

plot(mod.qp$fitted.values, rstandard(mod.qp))

```



Chap 6, Q4

(a)

- i. Each explanatory variable is assumed to have the same effect on each subject, but their overall (log) means (or intercepts; the “baselin” number of drinks when all explanatory variables are zero) are assumed to vary randomly.
- ii. We tried 1, 3, 5, 7, 9, 12, 15 quadrature points. The variance component estimate is 0.208 for 1 and 3, and 0.209 thereafter. Fixed effect parameter estimates are bouncing around seemingly randomly but not by more than the 3rd decimal place, so we conclude that the additional quadrature points beyond about 5 are not useful.

```
library(lme4)
data <- read.csv(
  "http://www.chrisbilder.com/categorical/Chapter4/DeHartSimplified.csv")
data[,3] = as.factor(data[,3])
mod <- glmer(formula = numall ~ nrel+ prel+ negevent+ posevent
  + gender+ ros+ age+ desired+ state + dayweek + (1|id),
  data = data, family = poisson(), nAGQ = 5)
mod.glm <- glm(formula = numall ~ dayweek + nrel+ prel + negevent
  +posevent+ gender+ ros+ age+ desired+ state
  + dayweek , data = data, family = poisson())
```

iii

$T = 43.744$  with p-value  $3.17e - 10$

```

m1 <- lmer(numall ~ dayweek + nrel+ prel
            + negevent+ posevent+ gender+ rosn
            + age+ desired+ state + (1|id), data = data,REML=F)
m0 <- lm(numall ~ dayweek + prel+ negevent
         + posevent+ gender+ rosn+ age
         + desired+ state, data = data,REML=F)
anova(m1,m0) ## two sequential tests
Data: data
Models:
m0: numall ~ dayweek + prel + negevent + posevent + gender + rosn +
m0:   age + desired + state
m1: numall ~ dayweek + nrel + prel + negevent + posevent + gender +
m1:   rosn + age + desired + state + (1 | id)
      Df    AIC    BIC logLik deviance  Chisq Chi Df Pr(>Chisq)
m0 16 2828.5 2899.3 -1398.3  2796.5
m1 18 2788.8 2868.4 -1376.4  2752.8 43.744    2  3.17e-10 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

iiii

	Df	AIC	LRT	Pr(Chi)
<none>		1034.85		
nrel	1.00	1040.49	7.65	0.01
prel	1.00	1033.24	0.39	0.53
negevent	1.00	1037.15	4.31	0.04
posevent	1.00	1032.85	0.00	0.97
gender	1.00	1033.41	0.56	0.45
rosn	1.00	1034.64	1.79	0.18
age	1.00	1033.19	0.34	0.56

```

scope = names(data)[-c(1,2,4)]
ltr1 = drop1(mod, scope = c("nrel","prel", "negevent", "posevent"),
            data = na.omit(data), test ="Chisq")
xtable(as.matrix(ltr2))
ltr2 = drop1(mod, scope = c("gender", "rosn", "age"),
            data = na.omit(data), test ="Chisq")

```

(b)

(i) Independence means  $y_{ij}$  and  $y_{ik}$  are independent; while exchangeable means that exchanging a pair of  $\mathbf{y}_i := \{y_{i1}, y_{i2}, \dots, y_{iJ}\}$ , the joint distribution of  $\mathbf{y}_i$  doesn't change.



(ii) There are significant effects due to `prel`, `negevent`, `desired`, and `dayweek`. The significance of `prel` is different from the GLMM, as is the nonsignificance of `nrel`.

```
library(geepack)
#data = data[sort(data[,1]),]
data = na.omit(data)
mod.gee <- geeglm(numall ~ nrel+ prel+ negevent+ posevent
                  + gender+ rosn+ age+ desired+ state + dayweek,
                  data = data, id = id, corstr = "exchangeable",
                  family = poisson("log"))

print(coef(summary(mod.gee)))
```

	Estimate	Std.err	Wald	Pr(> W )
(Intercept)	-0.326354009	0.74788112	0.19042009	6.625672e-01
nrel	0.090566890	0.04490711	4.06732368	4.372049e-02
prel	0.016480588	0.02780300	0.35136790	5.533398e-01
negevent	-0.291707311	0.13591679	4.60625943	3.185545e-02
posevent	0.013180937	0.06479952	0.04137605	8.388138e-01
gender	-0.119160707	0.11732462	1.03154412	3.097963e-01
rosn	0.129808682	0.12798412	1.02871563	3.104605e-01
age	-0.004888138	0.01110577	0.19372666	6.598329e-01
desired	0.278878955	0.03947359	49.91351724	1.606715e-12
state	-0.078026059	0.08945524	0.76079522	3.830798e-01
dayweek2	-0.177600673	0.13547804	1.71850742	1.898853e-01
dayweek3	-0.115100268	0.11314645	1.03483425	3.090259e-01
dayweek4	0.094241360	0.10979824	0.73670288	3.907191e-01
dayweek5	0.228328975	0.11068685	4.25529854	3.912807e-02
dayweek6	0.534209881	0.12147080	19.34105104	1.093304e-05
dayweek7	0.242976747	0.14411676	2.84249981	9.180129e-02

(iii) The test results change because the tests are performed assuming that variables are entered into the model in the order in which they are listed in the formula. Now `negevent` and `prel` are no longer significant, but `nrel` is.

(iiii) From the Anova table below, `dayweek` is significant.

	Df	AIC	BIC	logLik	deviance	Chisq	Chi Df	Pr(>Chisq)
mod.wo.dayweek	11.00	1103.47	1152.16	-540.73	1081.47			
mod	17.00	1034.85	1110.10	-500.42	1000.85	80.62	6.00	0.00

```
mod <- glmer(formula = numall~ nrel+ prel+ negevent+ posevent+ gender
+ rosn+ age+ desired+ state + dayweek + (1|id),
data = data, family = poisson(), nAGQ = 5)
```

```
mod.wo.dayweek <- glmer(formula = numall~nrel+ prel+ negevent+ poseven+ gender
+ rosn+ age+ desired+ state + (1|id),
data = data, family = poisson(), nAGQ = 5)
```

```
a = anova(mod,mod.wo.dayweek,test = "Chisq")
```